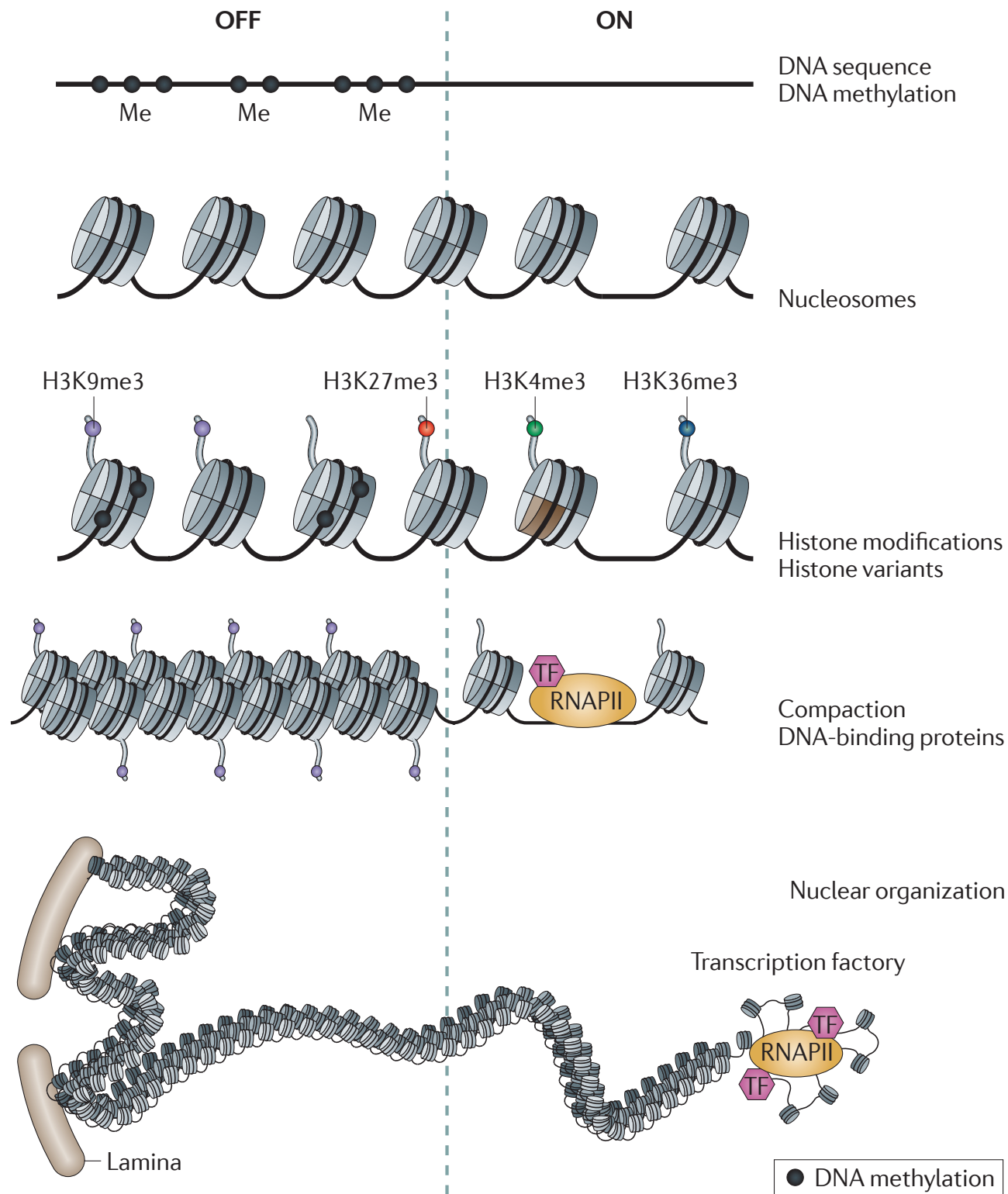


applied genomics

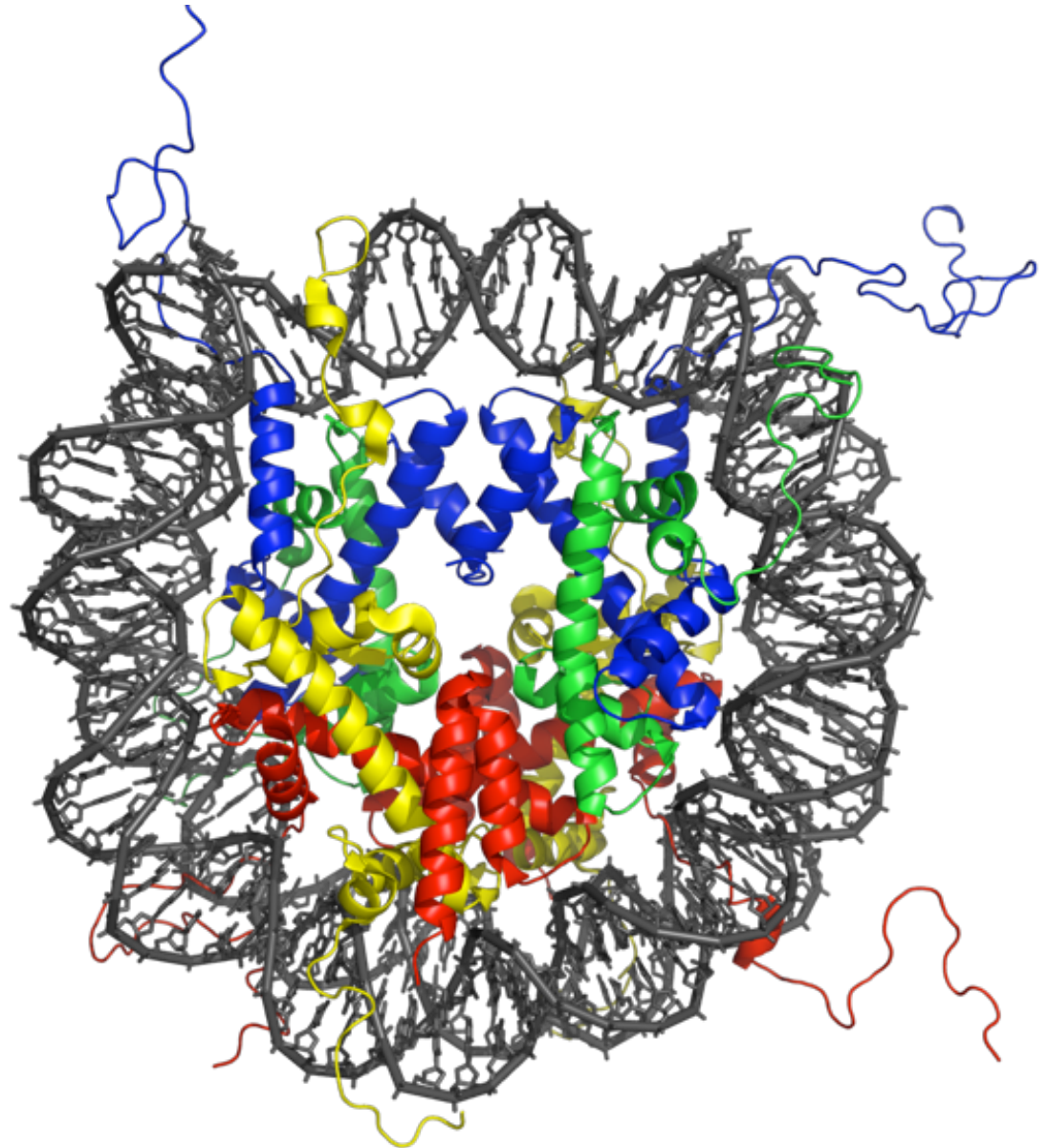
BIOL5V01 - Applied Genomics Laboratory Course

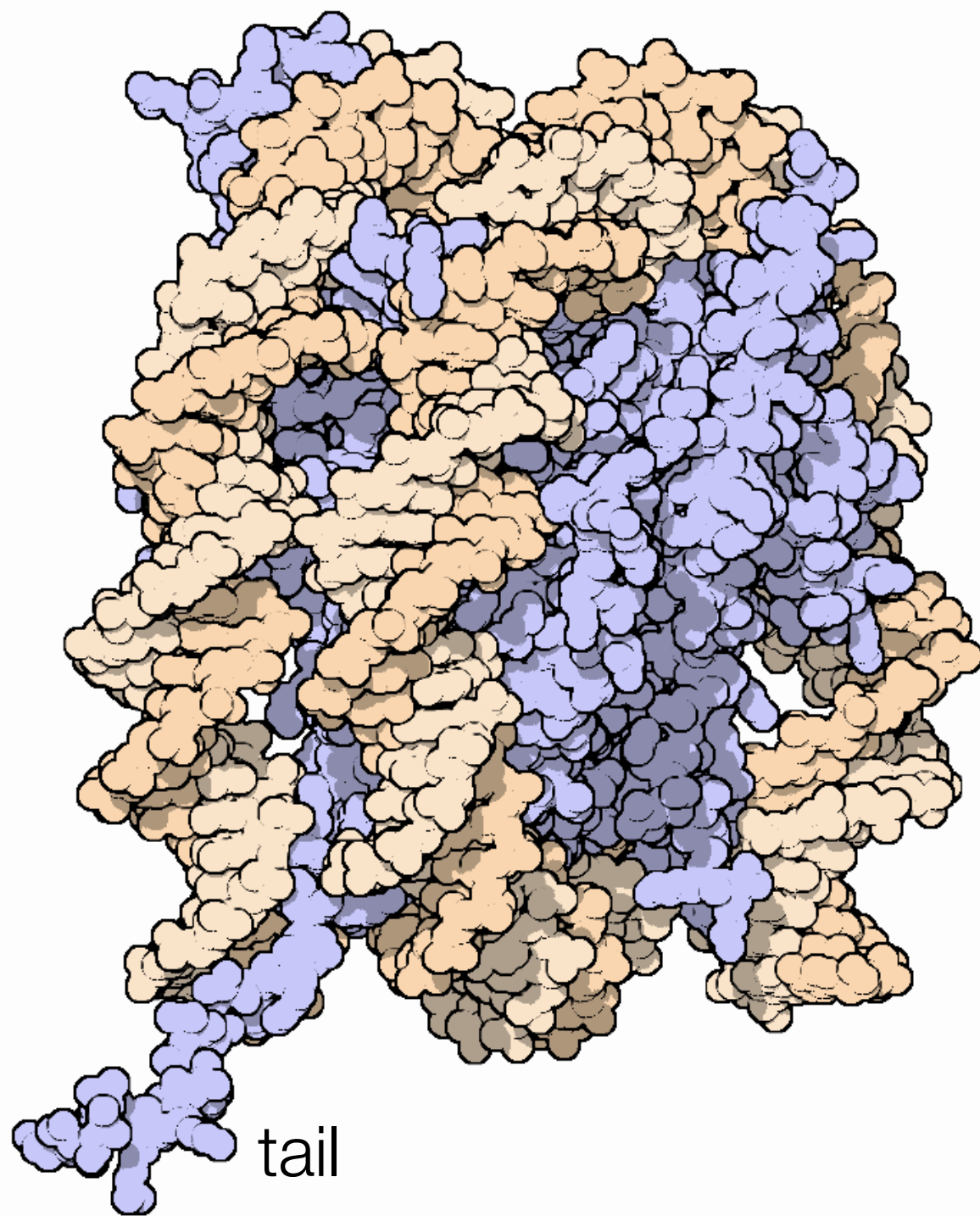
*Tae Hoon Kim, Ph.D.
genome@utdallas.edu
<http://taehoonkim.org>*



nucleosome - substrate for genetic processes

- *equal mass of histone to DNA in nucleus*
- *146bp DNA*
- *histones **H2A** , **H2B** , **H3** and **H4***
- *histone tails extend out from the nucleosome*
- *histone tails can be covalently modified at a number of specific residues*
- *nucleosomes can slide along the DNA or disassemble by ATP-dependent remodeling*
- *histone variants: H2AZ, H2AX, macroH2A, H3.3, CENPA*
- *linker histones*



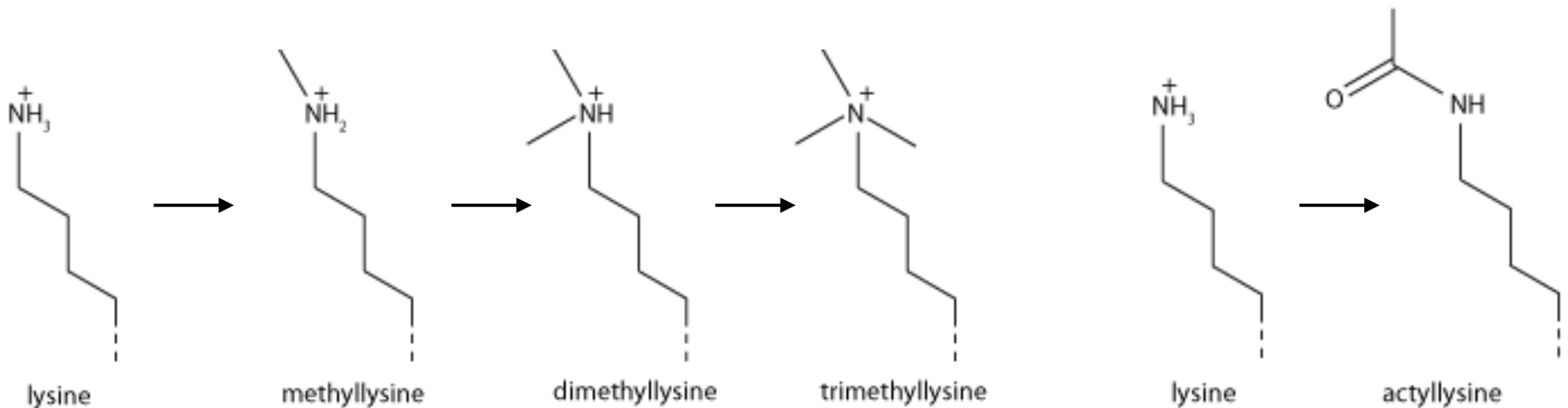


histone code

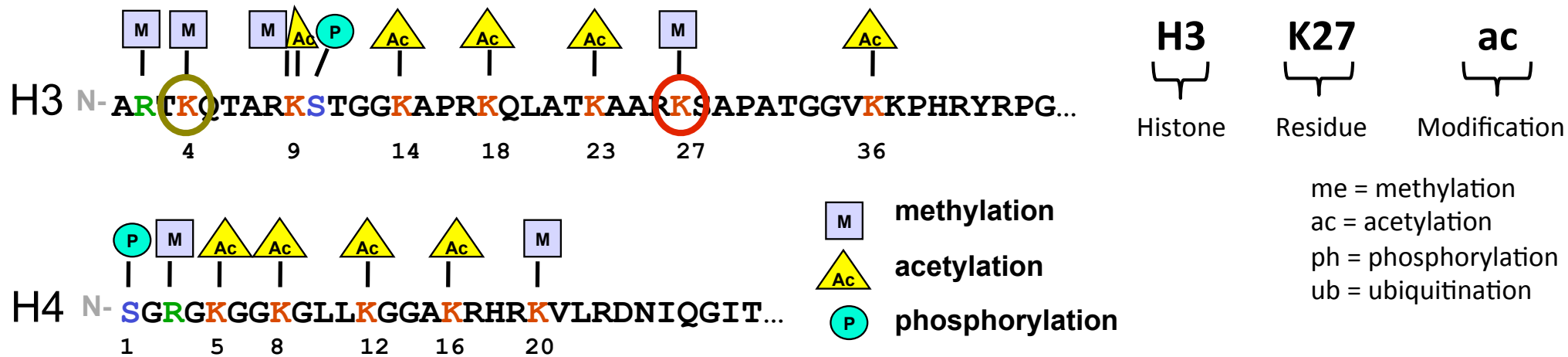
- *histones are an active component of epigenome*
- *histones compose equal amount of mass as DNA in the nucleus*
- *histone modifications single or combination serve as a code for regulation*
- *writers and readers of histone code*
 - *histone modifying enzymes are writers*
 - *modified histone binders are readers*

histone modifications and their functional associations

Type of modification	H3K4	H3K9	H3K14	H3K27	H3K79	H3K36	H4K20	H2BK5
mono-methylation	+	+		+	+		+	+
di-methylation		-		-	+			
tri-methylation	+	-		-	+, -	+		-
acetylation		+	+	+				



histone code

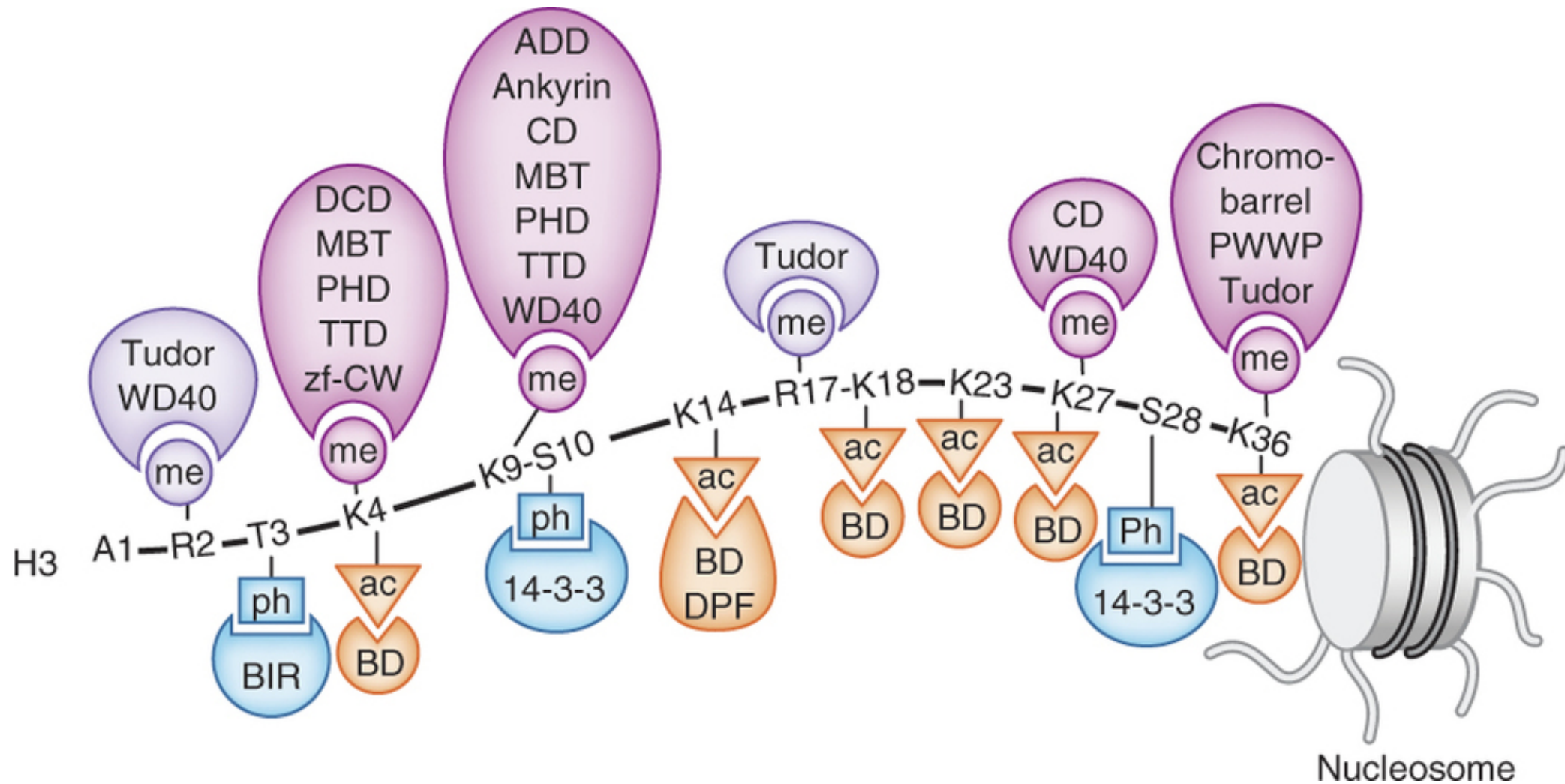


- specific histone modifications serve as a docking site for other regulatory proteins that reads the marks
- specific histone modifications indicate gene activity and chromatin structures:
 - trimethylation of lysine **4** on histone H3 (H3K4me3) is associated with active genes (Trithorax)
 - trimethylation of lysine **27** on histone H3 (H3K27me3) is associated with silent genes (Polycomb)
- histone modifications are reversible and dynamic:
 - acetylated histones can be deacetylated
 - methylated histones can be demethylated

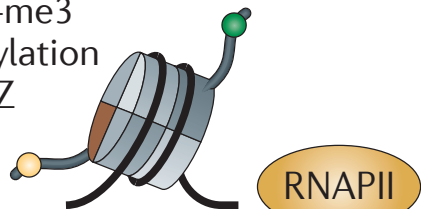
histone code writers/erasers

- *histone acetyl transferases (HAT)*
- *histone deacetylases (HDAC)*
- *lysine methyl transferases (KMT)*
- *lysine demethylases (KDM)*
- *kinases*
- *ubiquitin ligases*
- *deubiquitinating enzymes*
- *poly-ADP-ribose polymerase (PARP)*
- *poly-ADP-ribose glycohydrolase (PARG)*
- *...*

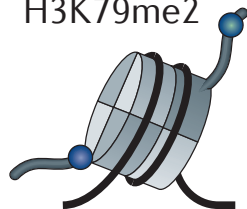
histone code readers



H3K4me2
H3K4me3
Acetylation
H2A.Z



H3K36me3
H3K79me2



RNAPII

Exon

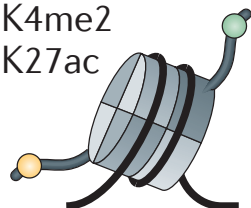
Intron

Exon

p300

Enhancer

H3K4me1
H3K4me2
H3K27ac



CTCF

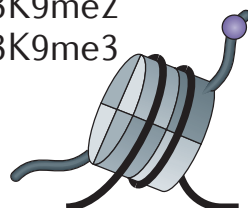
Boundary

Exon

Intron

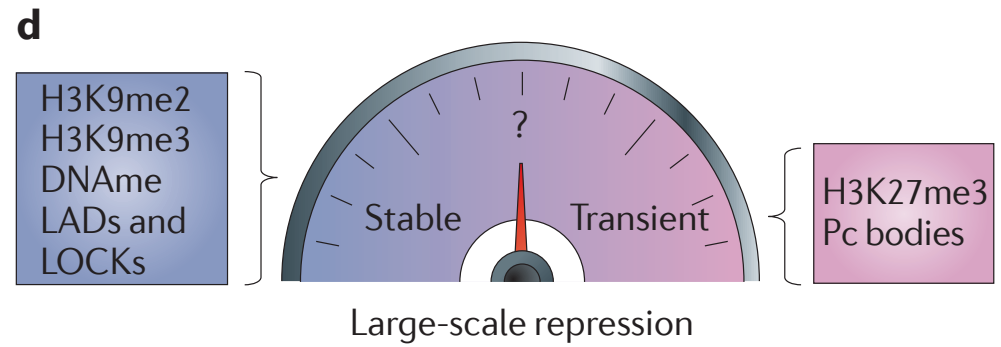
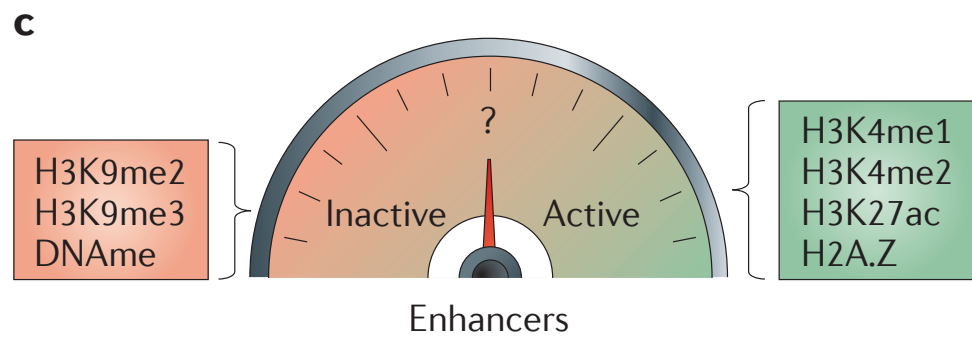
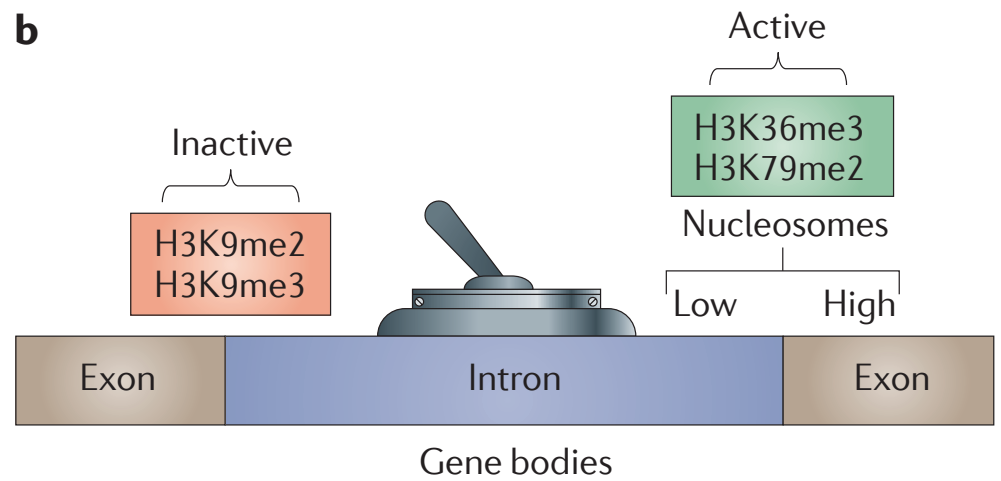
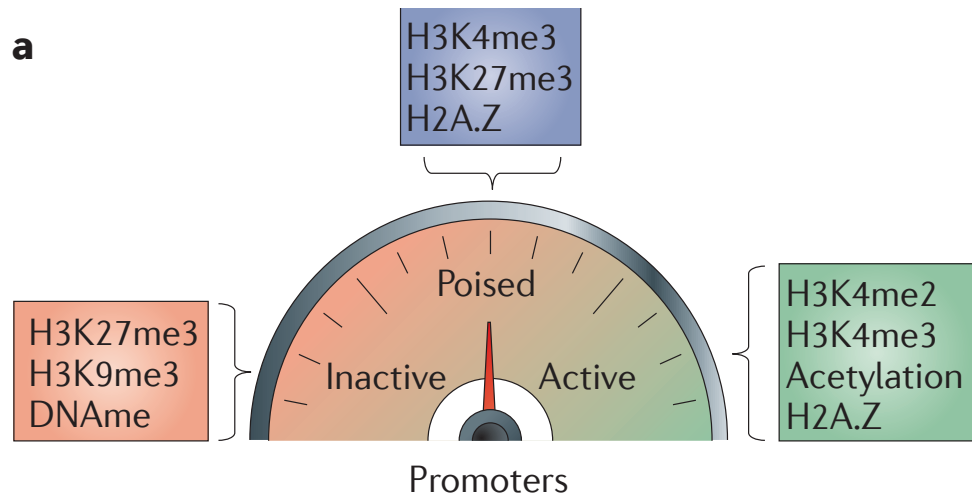
Exon

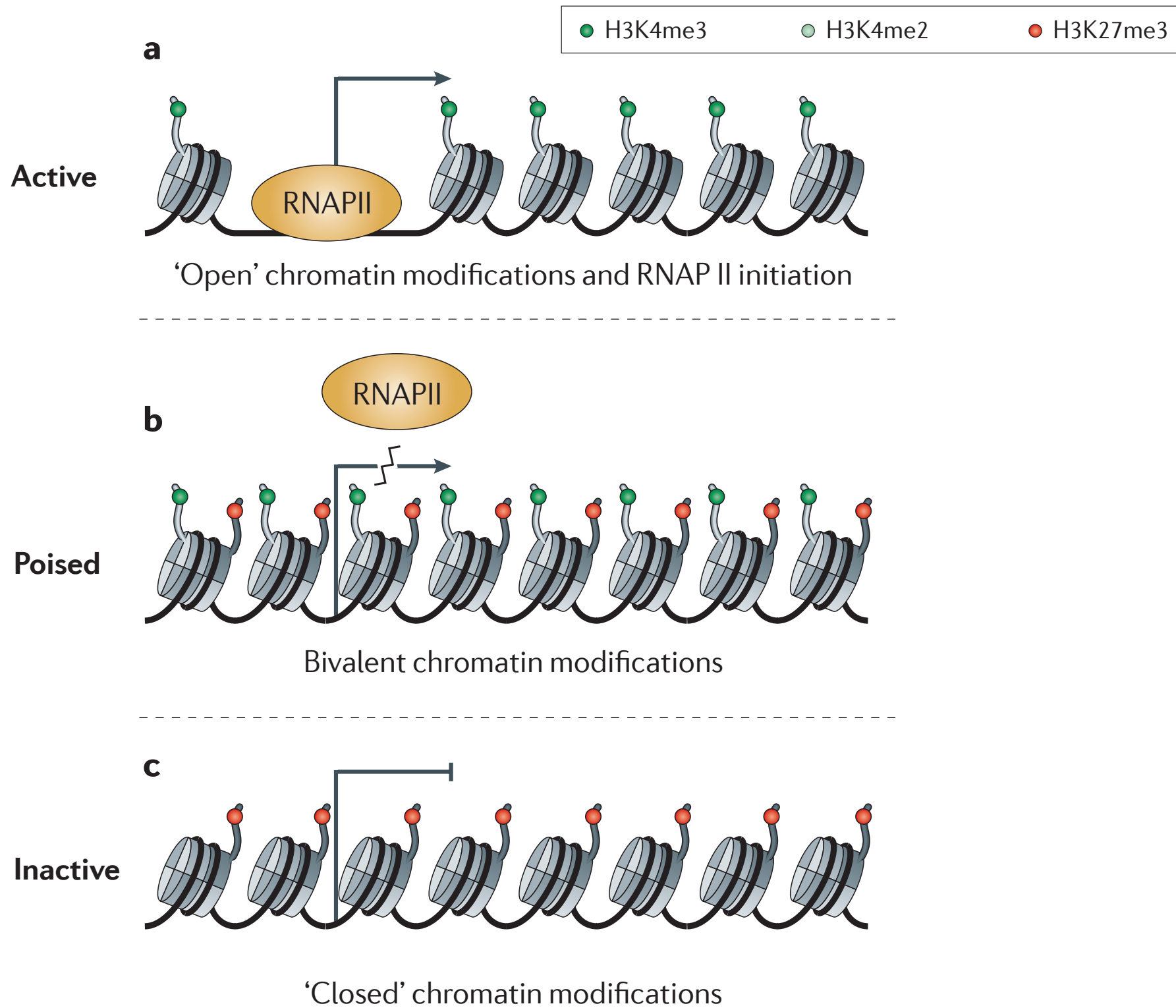
H3K9me2
H3K9me3

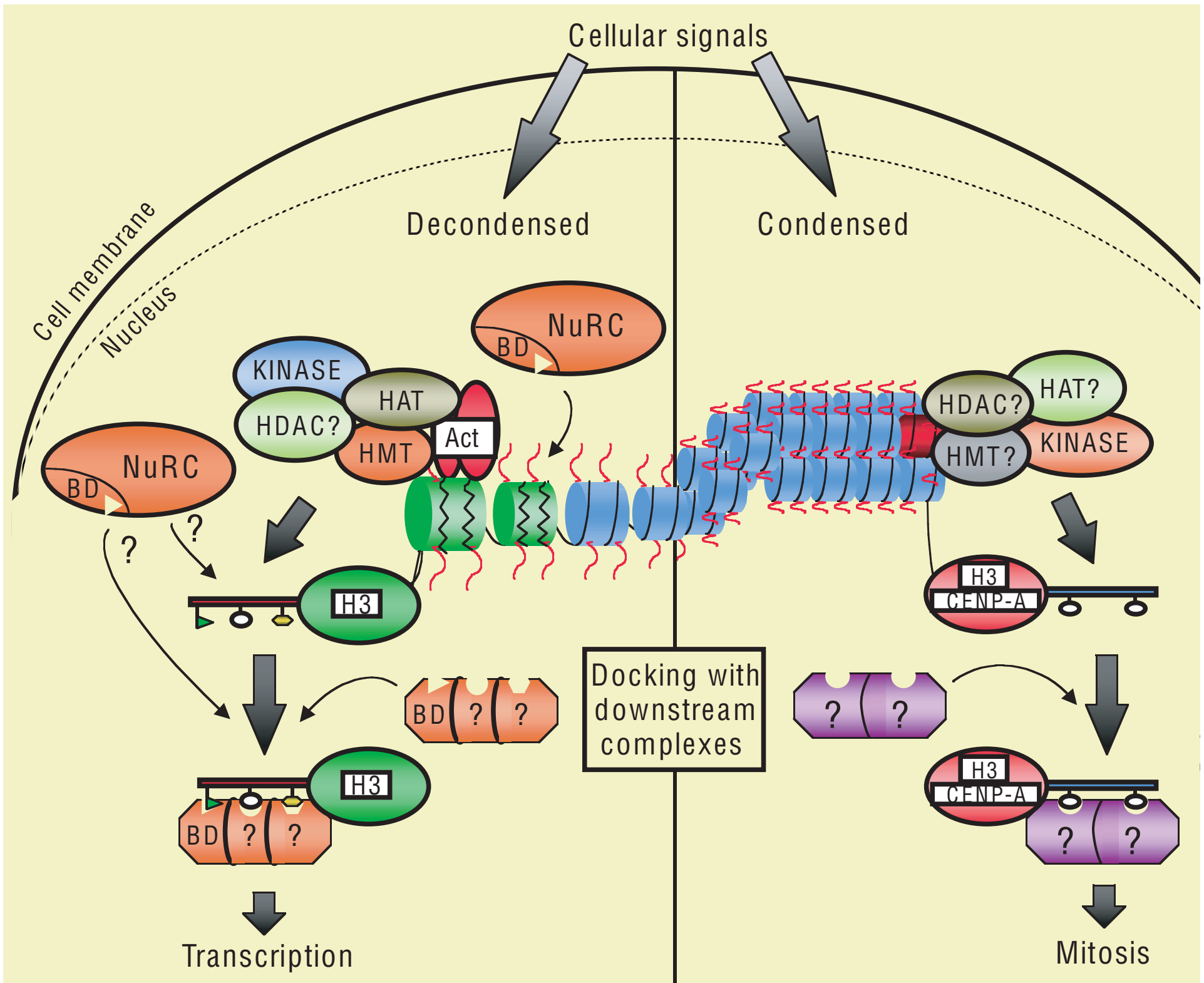


H3K27me3

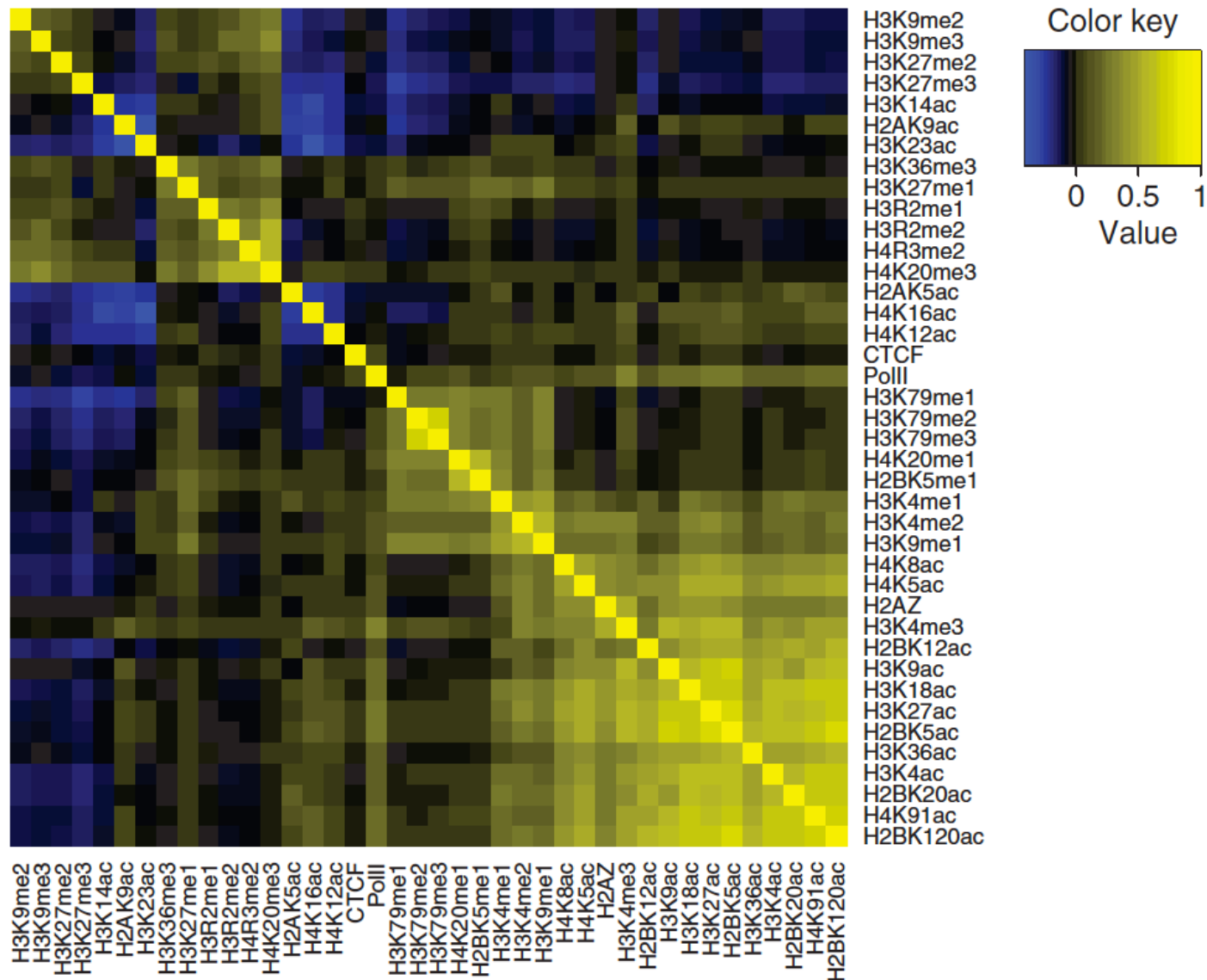




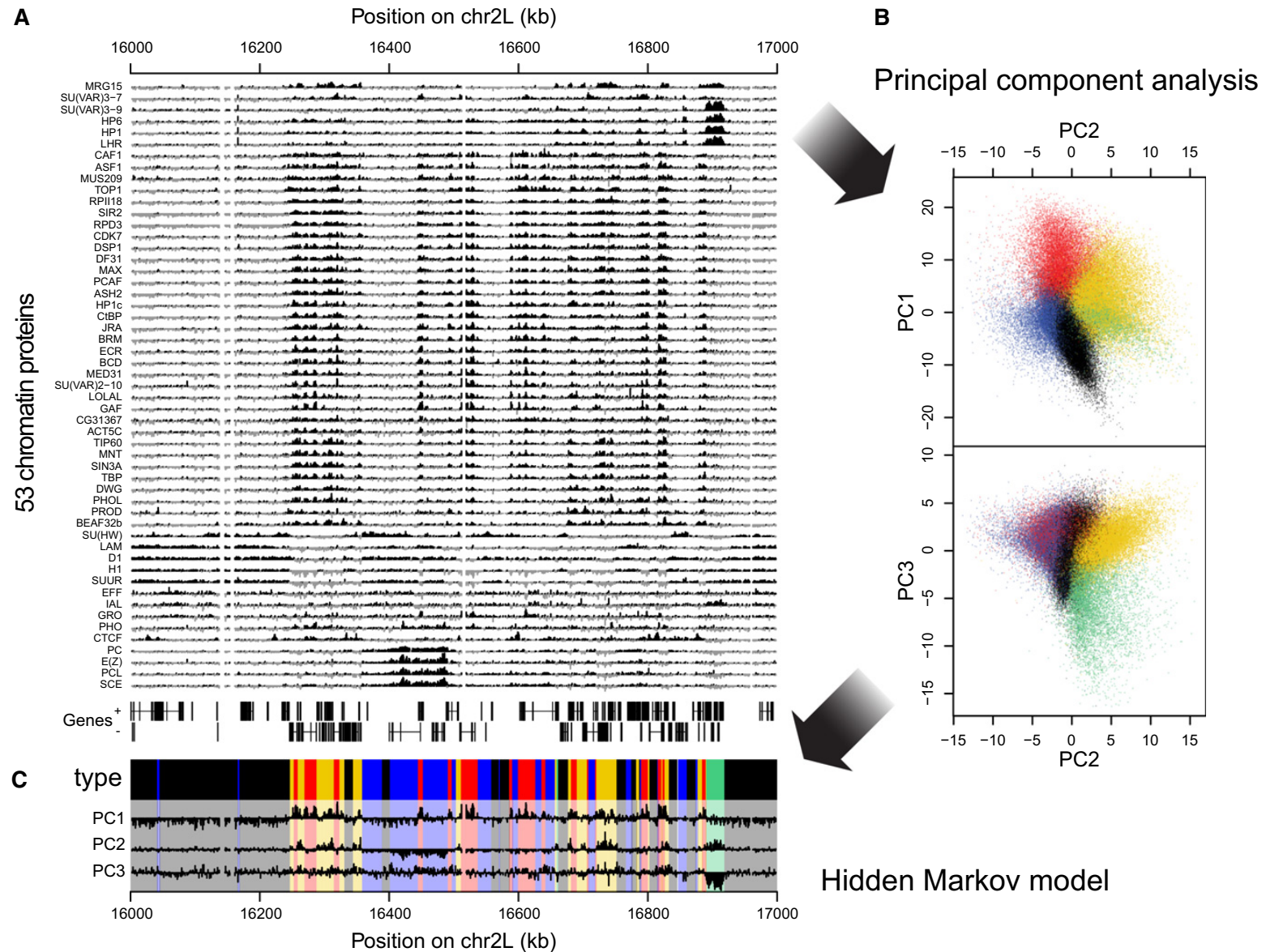




histone modifications are correlated



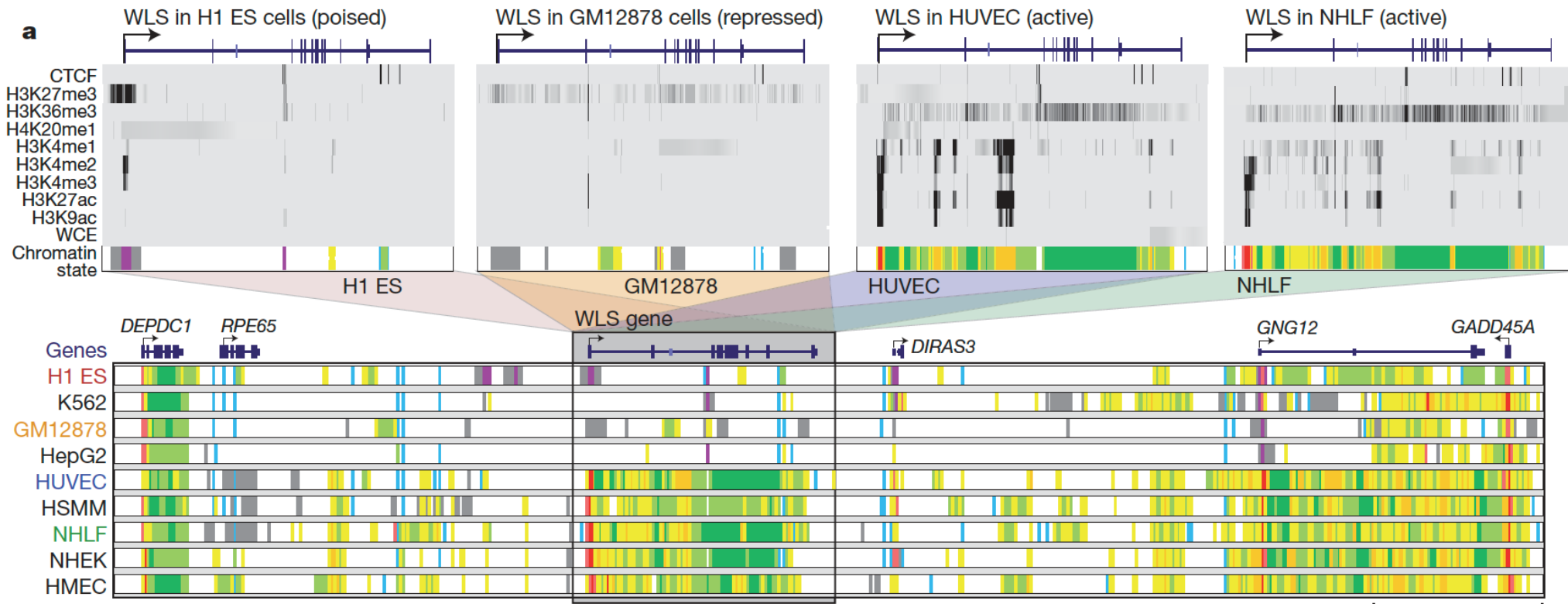
five colors of Drosophila chromatin



modeling chromatin states in the human genome

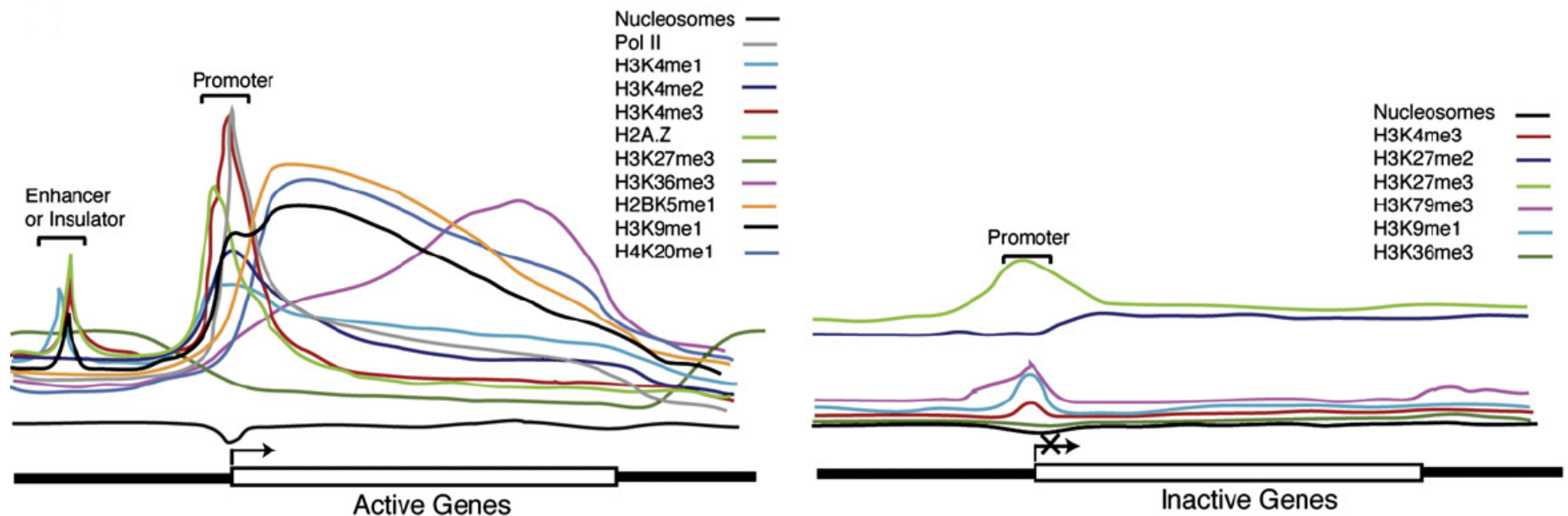
Chromatin states	State	Chromatin mark observation frequency (%)										Functional enrichments (fold)							Candidate state annotation
		CTCF	H3K27me3	H3K36me3	H4K20me1	H3K4me1	H3K4me2	H3K4me3	H3K27ac	H3K9ac	WCE	±2 kb TSS	Conserved non-exon	DNase (K562)	c-Myc (K562)	NF-κB (GM12878)	Transcript	Nuclear lamina (NHLF)	
	1	16	2	2	6	17	93	99	96	98	2	83	3.8	23.3	82.0	40.7	0.2	0.15	Active promoter
	2	12	2	6	9	53	94	95	14	44	1	58	2.8	15.3	12.6	5.8	0.6	0.30	Weak promoter
	3	13	72	0	9	48	78	49	1	10	1	49	4.3	10.8	3.1	1.0	0.4	0.68	Inactive/poised promoter
	4	11	1	15	11	96	99	75	97	86	4	23	2.7	23.1	31.8	49.0	1.3	0.05	Strong enhancer
	5	5	0	10	3	88	57	5	84	25	1	3	1.8	13.6	6.3	15.8	1.4	0.10	Strong enhancer
	6	7	1	1	3	58	75	8	6	5	1	17	2.4	11.9	5.7	7.0	1.1	0.31	Weak/poised enhancer
	7	2	1	2	1	56	3	0	6	2	1	4	1.5	5.1	0.6	2.4	1.3	0.20	Weak/poised enhancer
	8	92	2	1	3	6	3	0	0	1	1	3	1.5	12.8	2.5	1.2	1.1	0.61	Insulator
	9	5	0	43	43	37	11	2	9	4	1	4	1.1	4.5	0.7	0.8	2.4	0.02	Transcriptional transition
	10	1	0	47	3	0	0	0	0	0	1	1	0.9	0.3	0.0	0.0	2.5	0.11	Transcriptional elongation
	11	0	0	3	2	0	0	0	0	0	0	2	0.9	0.3	0.0	0.1	1.9	0.24	Weak transcribed
	12	1	27	0	2	0	0	0	0	0	0	5	1.4	0.3	0.0	0.1	0.8	0.63	Polycomb repressed
	13	0	0	0	0	0	0	0	0	0	0	1	0.9	0.1	0.0	0.0	0.7	1.30	Heterochrom; low signal
	14	22	28	19	41	6	5	26	5	13	37	3	0.4	1.9	0.3	0.2	0.4	1.44	Repetitive/CNV
	15	85	85	91	88	76	77	91	73	85	78	1	0.2	5.9	9.5	7.4	0.4	1.30	Repetitive/CNV

chromatin states vary across cell types



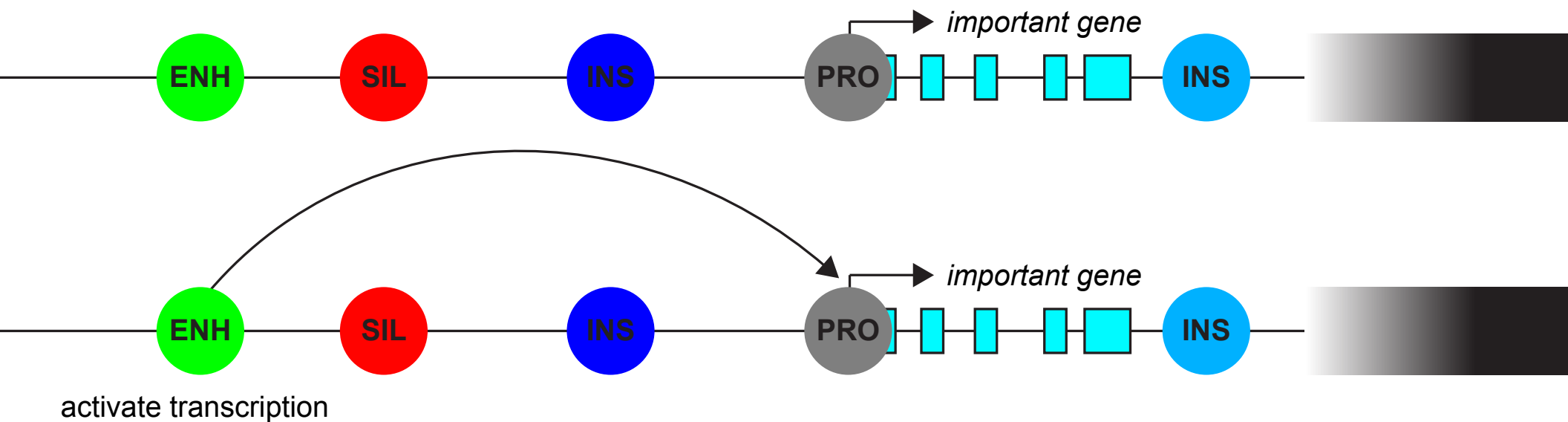
histone methylation at promoters, enhancers and gene bodies

T-cells

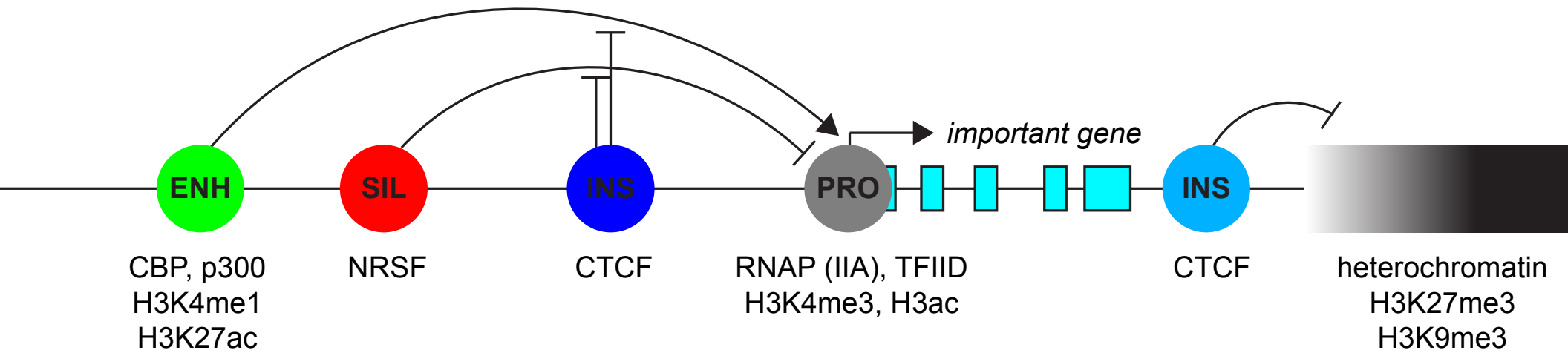


- *gene activity and histone modification*
 - *genes activity can be predicted from histone modification signature*
 - *direction of transcription can be determined from histone modification pattern*
- *RNA polymerase is paused at the promoter - transcription elongation (not polymerase recruitment) might be the rate limiting step for gene expression*

how do you regulate a gene?

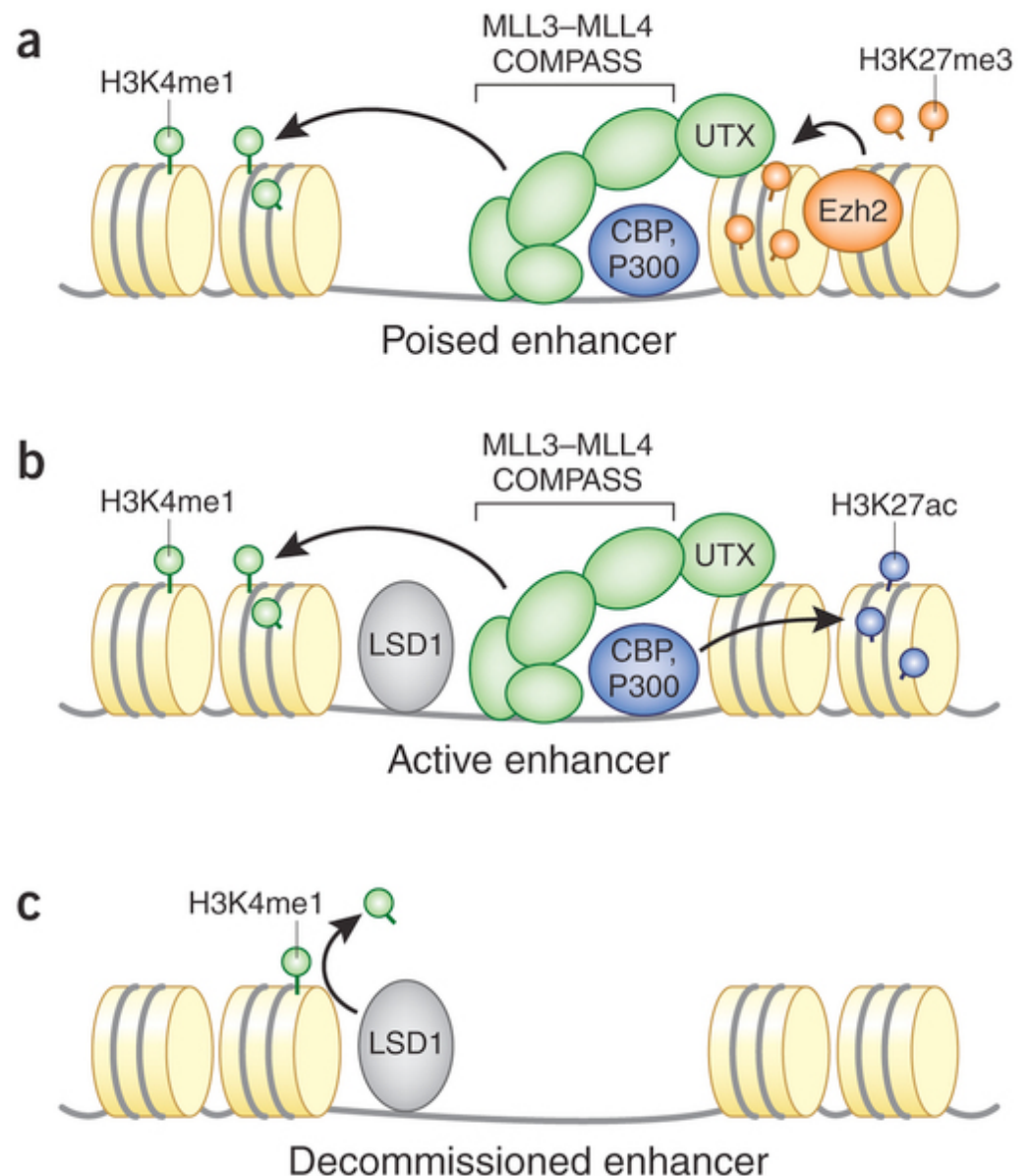


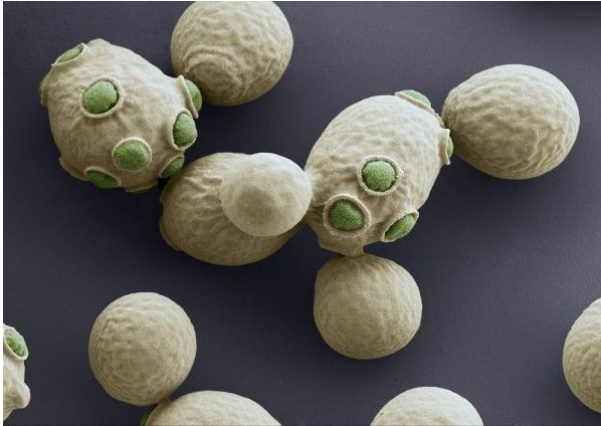
- *sequence specific transcription factors bind to enhancer and promoter to recruit the transcription machinery (general transcription factors and RNA polymerase) to synthesize mRNA*
- *how to specify regulation?*
 - *combinatorial regulation*
 - *coregulators (co-activators/co-repressor) serve to integrate cis-regulatory signal*



enhancer regulation

- *poised enhancers are marked with H3K4me1 by MLL3/4 (TrxG) complex*
- *active enhancer recruits p300/CBP and acetylates histones resulting in H3K27ac*
- *when enhancer is decommissioned, histone demethylase LSD1 is recruited to remove H3K4me1*



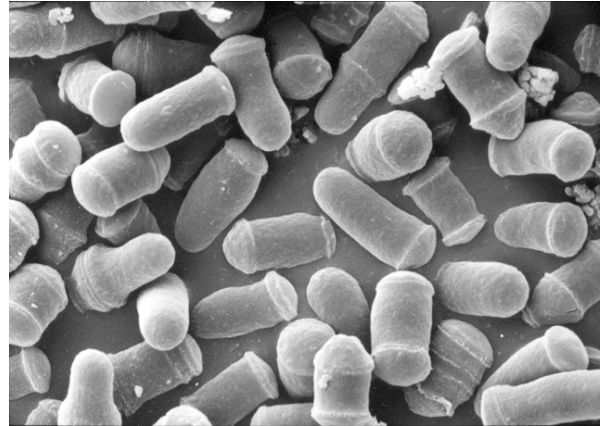


budding yeast

- 12 chromosomes
- 12Mb
- no HMT for H3K9
- no HP1 homolog

yeastgenome.org

unmodified chromatin



fission yeast

- 3 large chromosomes
- 12.6 Mb
- many repeats and transposons
- H3K9me3
- HP1 homolog

pombase.org

H3K9me3



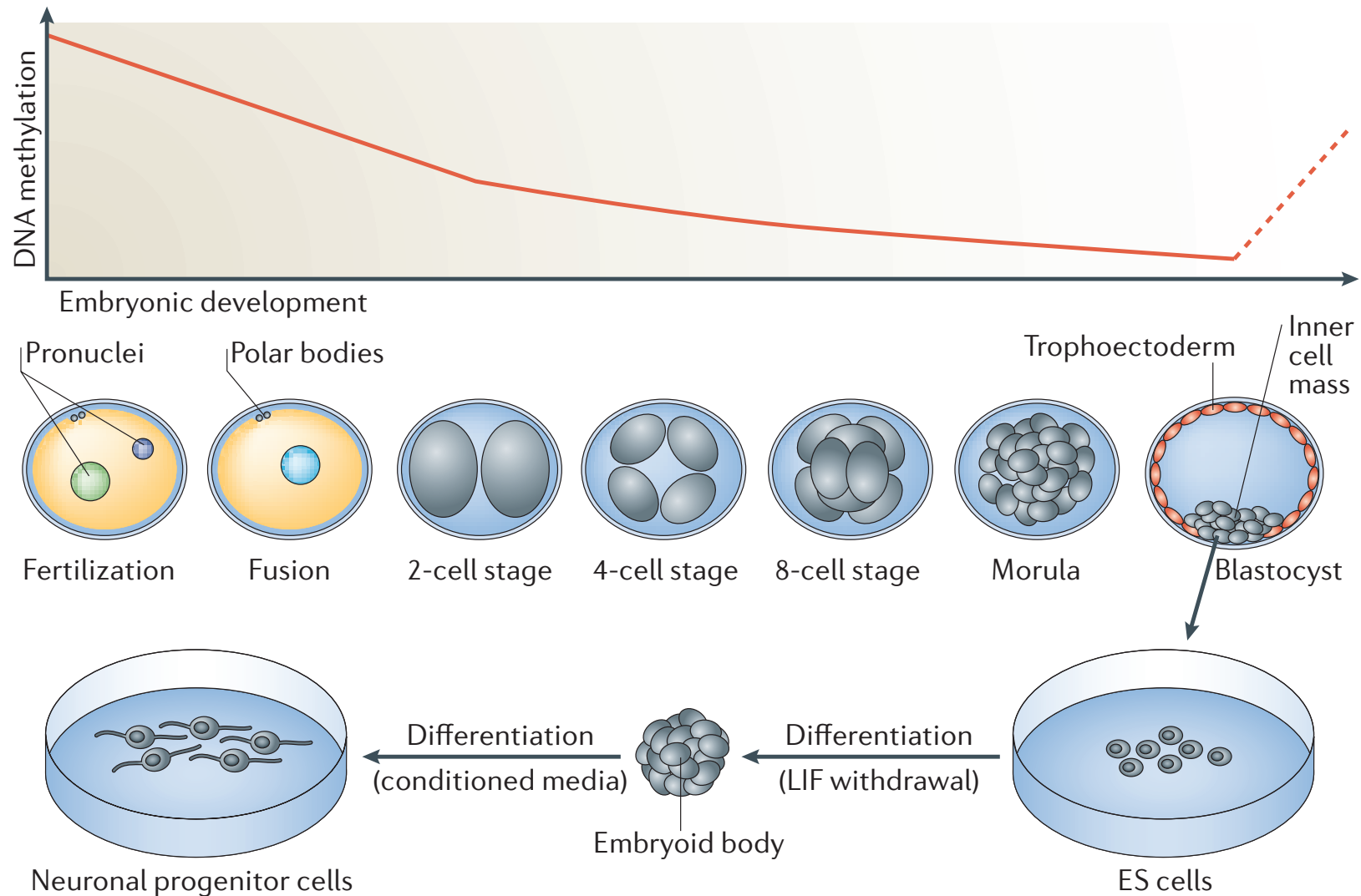
fruit fly

- 5 chromosomes
- 118.4Mb
- many repeats and transposons
- H3K9me3
- HP1
- H3K27me3
- Polycomb

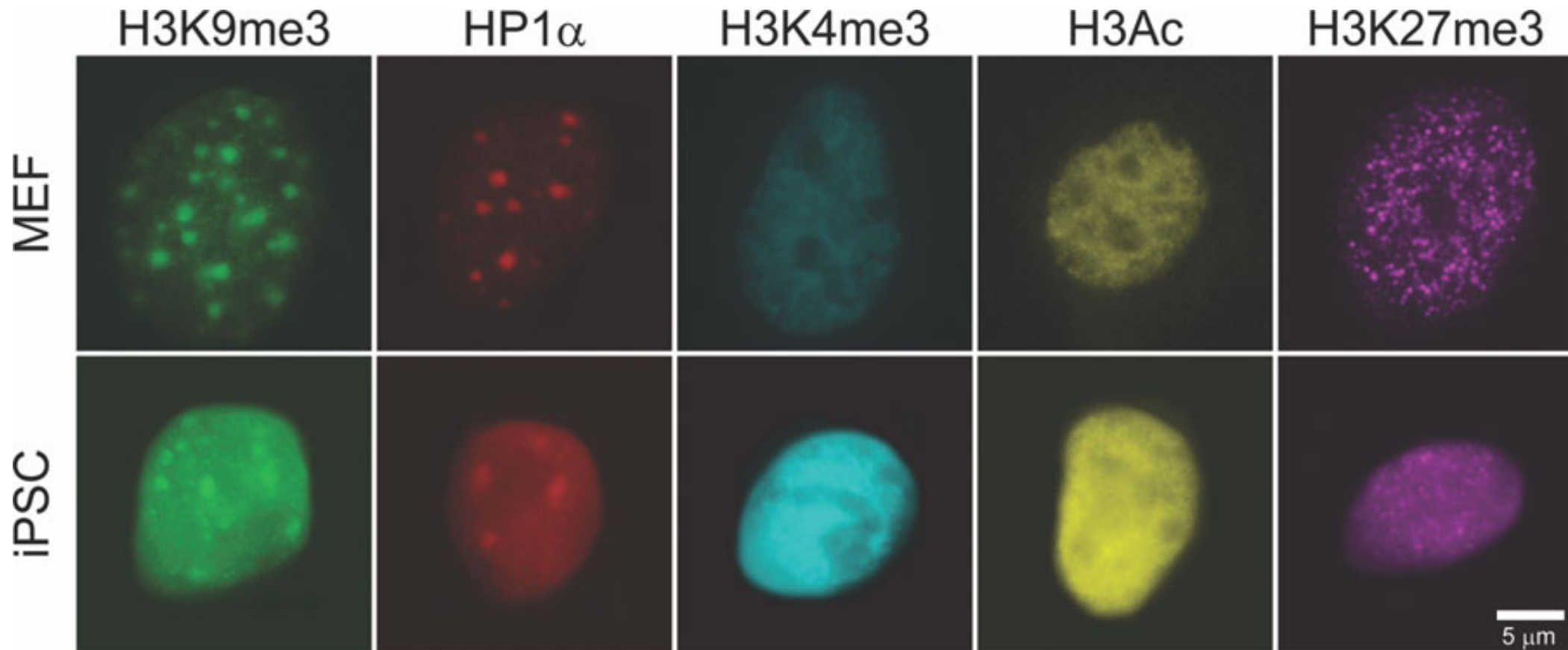
flybase.org

H3K9me3, H3K27me3

DNA methylation changes during development



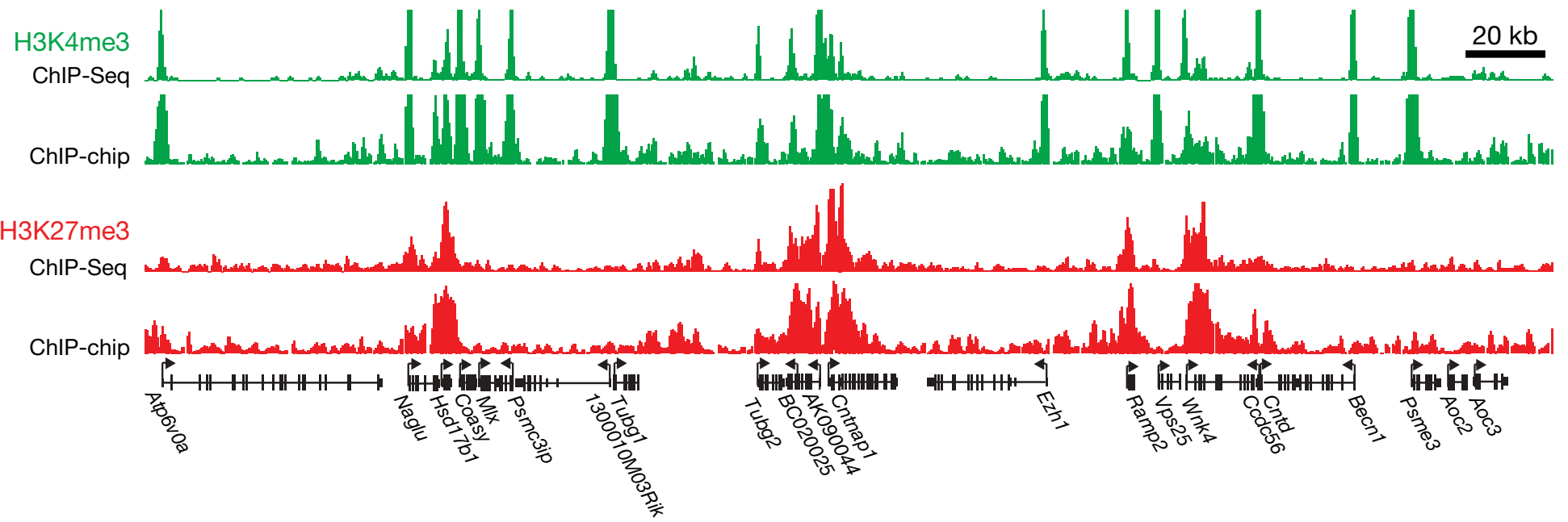
nuclear differences between embryonic stem cells and differentiated cells

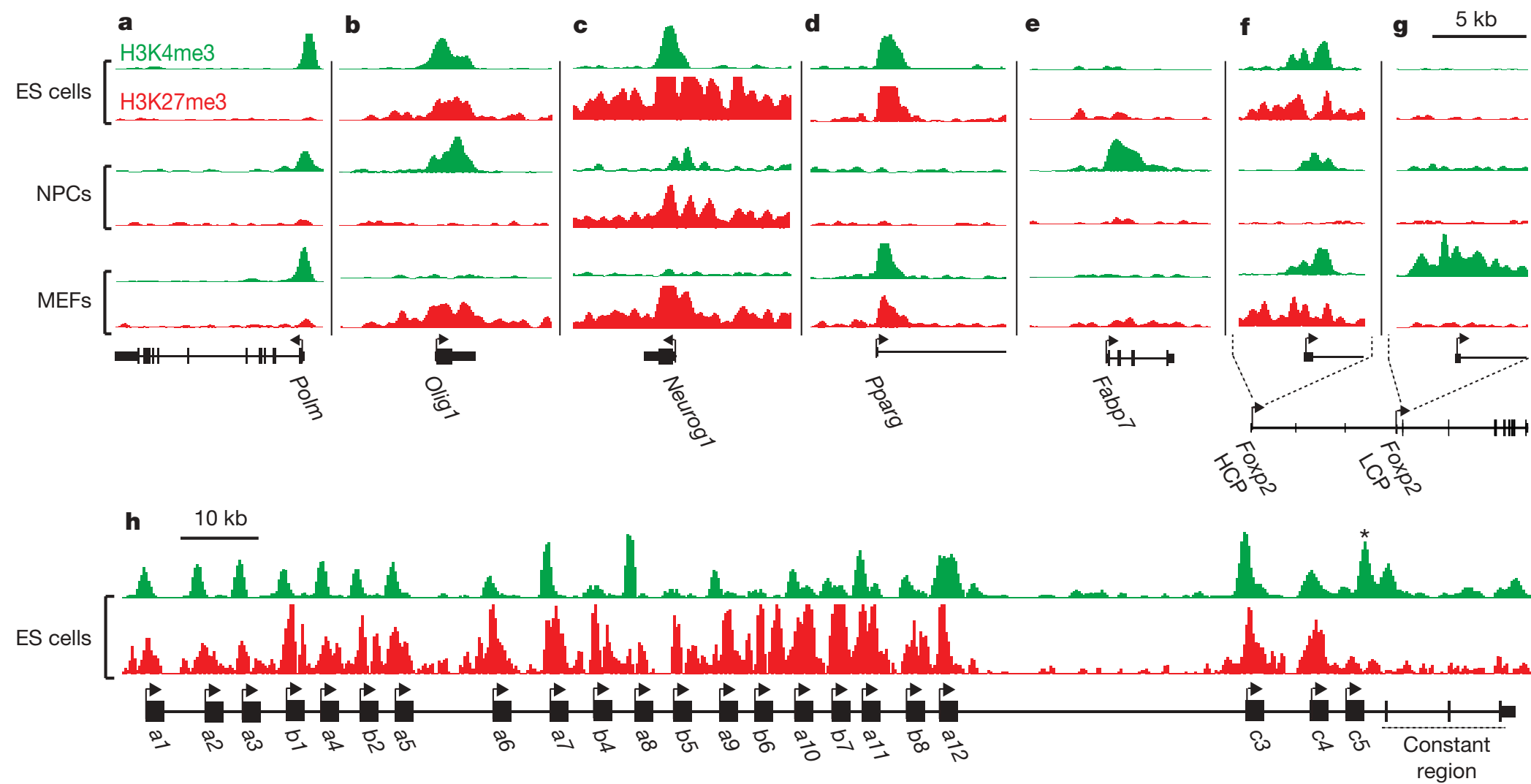


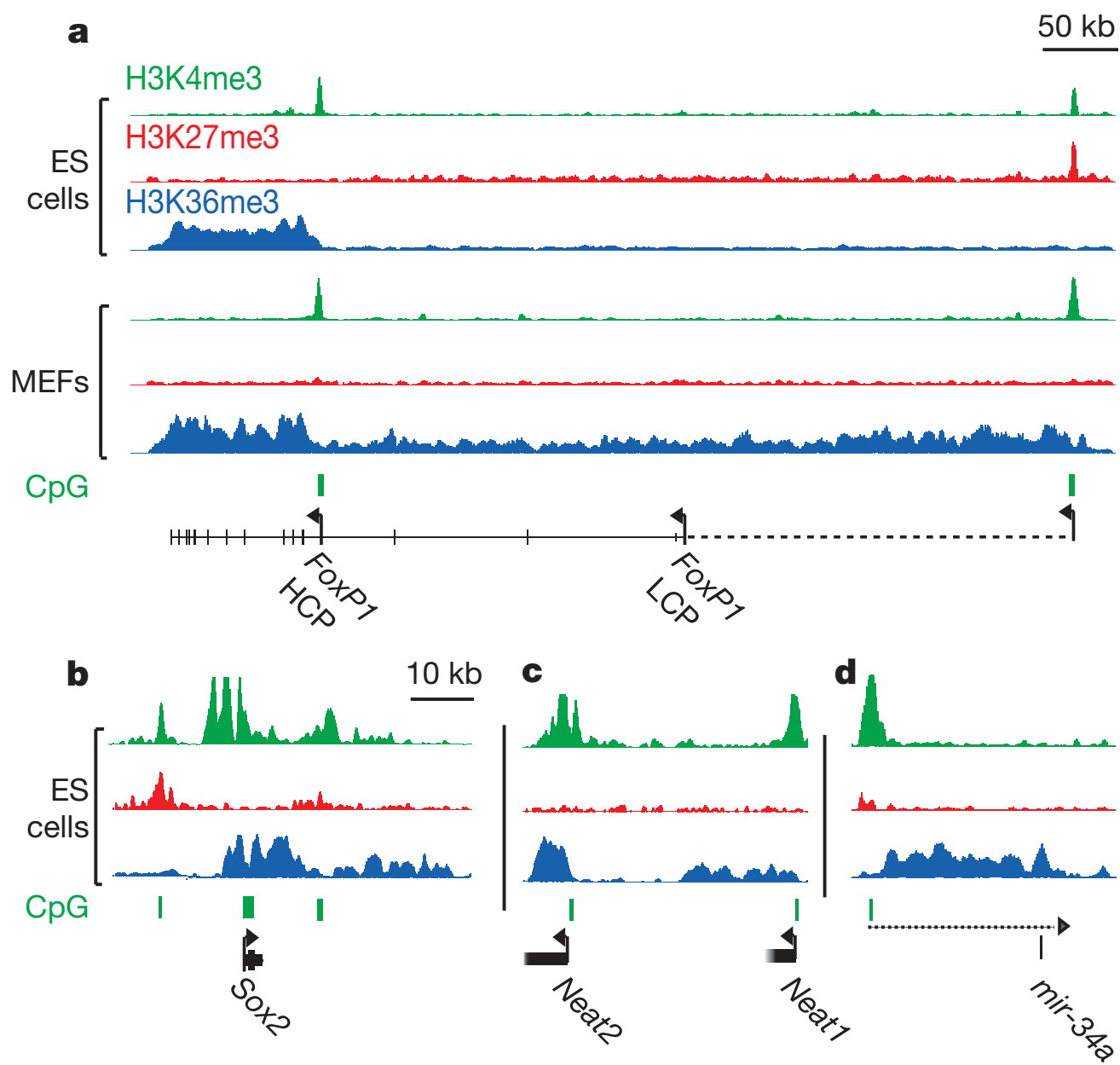
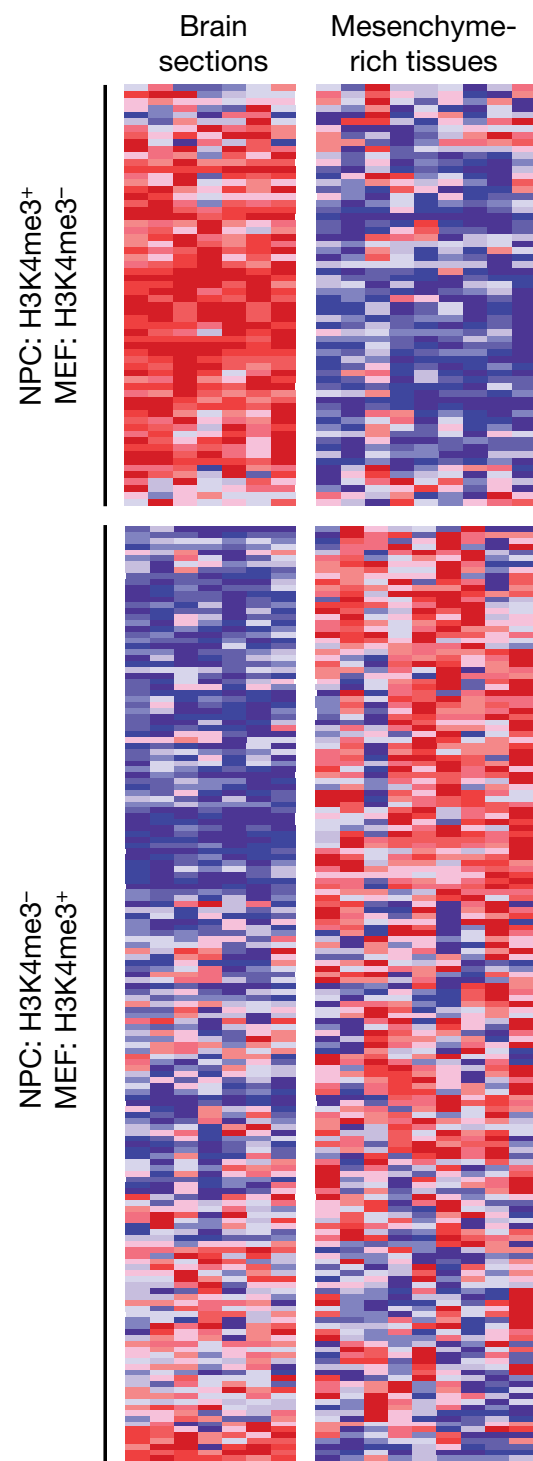
ARTICLES

Genome-wide maps of chromatin state in pluripotent and lineage-committed cells

Tarjei S. Mikkelsen^{1,2}, Manching Ku^{1,4}, David B. Jaffe¹, Biju Issac^{1,4}, Erez Lieberman^{1,2}, Georgia Giannoukos¹, Pablo Alvarez¹, William Brockman¹, Tae-Kyung Kim⁵, Richard P. Koche^{1,2,4}, William Lee¹, Eric Mendenhall^{1,4}, Aisling O'Donovan⁴, Aviva Presser¹, Carsten Russ¹, Xiaohui Xie¹, Alexander Meissner³, Marius Wernig³, Rudolf Jaenisch³, Chad Nusbaum¹, Eric S. Lander^{1,3*} & Bradley E. Bernstein^{1,4,6*}







Encyclopedia of DNA
elements (ENCODE)
Project since 2003



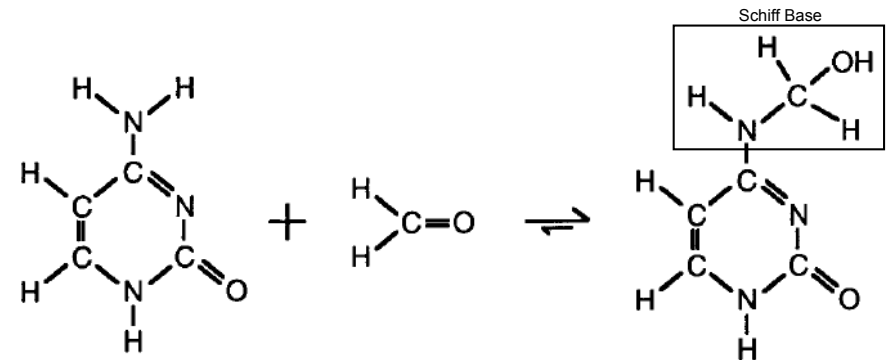
<http://www.nature.com/encode/#/threads>

<http://genome.ucsc.edu/ENCODE>

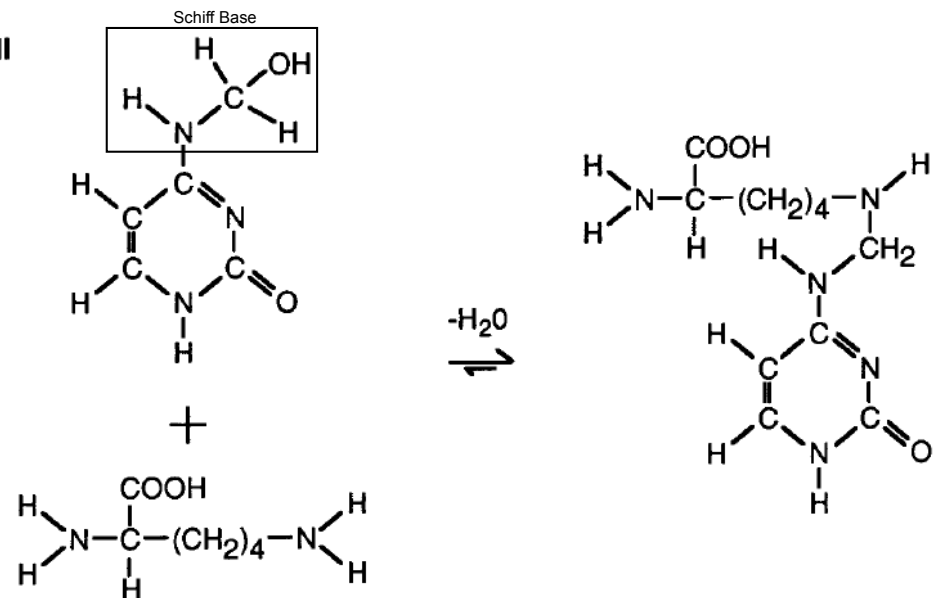
chemistry of formaldehyde crosslinking

- *versatile 2 Å crosslinker*
- *highly cell permeable*
- *limited by availability of primary amines in the vicinity*
- *a selective crosslinker - not general*
 - *requires close proximity of free amines*
 - *some proteins not crosslinked*
- *crosslinks protein-proteins, protein-DNA contacts*
- *reversible*

Reaction I



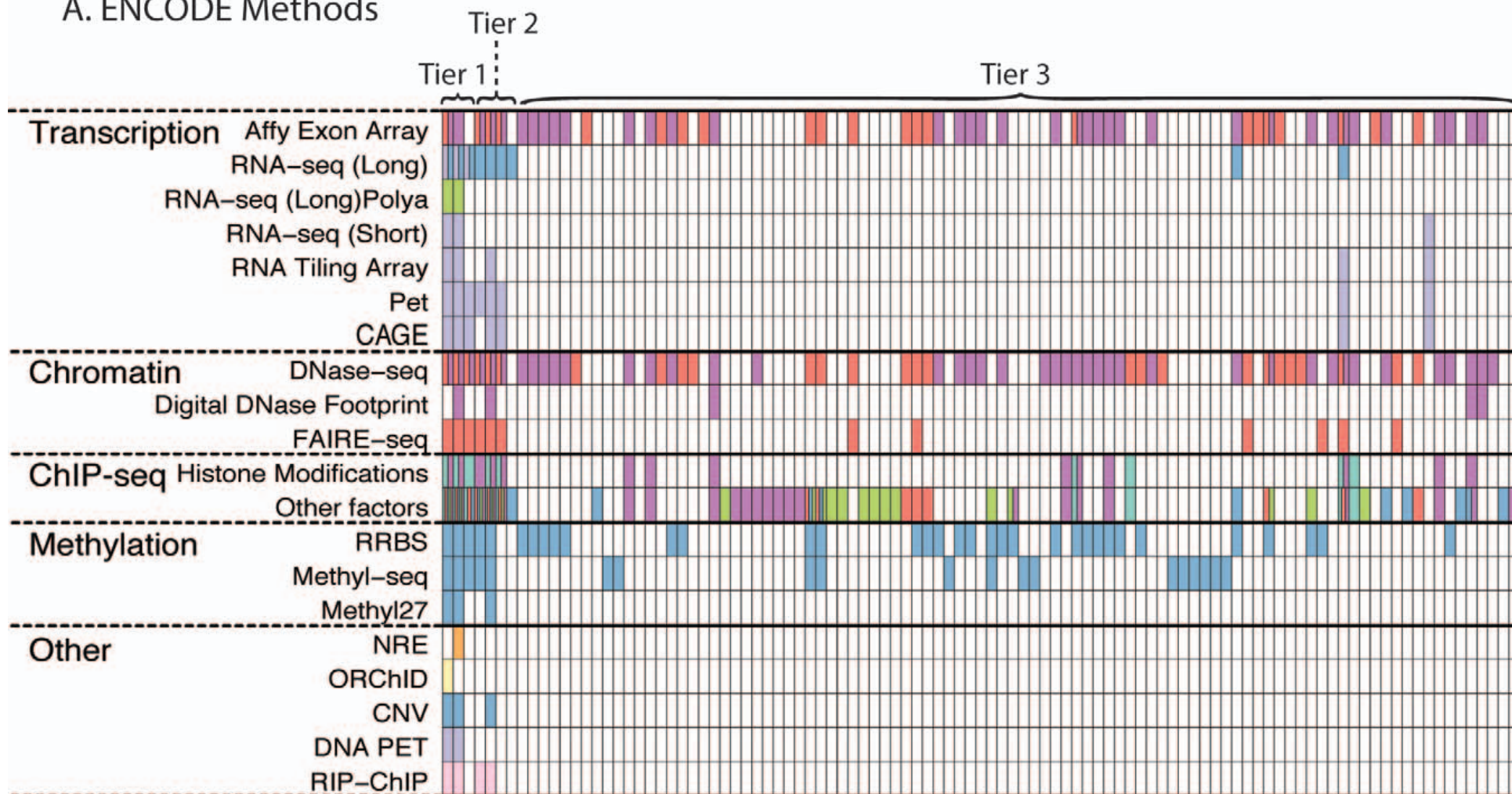
Reaction II



considerations

- *direct vs indirect associations*
- *ensemble measurement of association*
 - *can't discern binding at individual chromosomes*
- *requires specific antibodies*
- *tagging not always effective*

A. ENCODE Methods



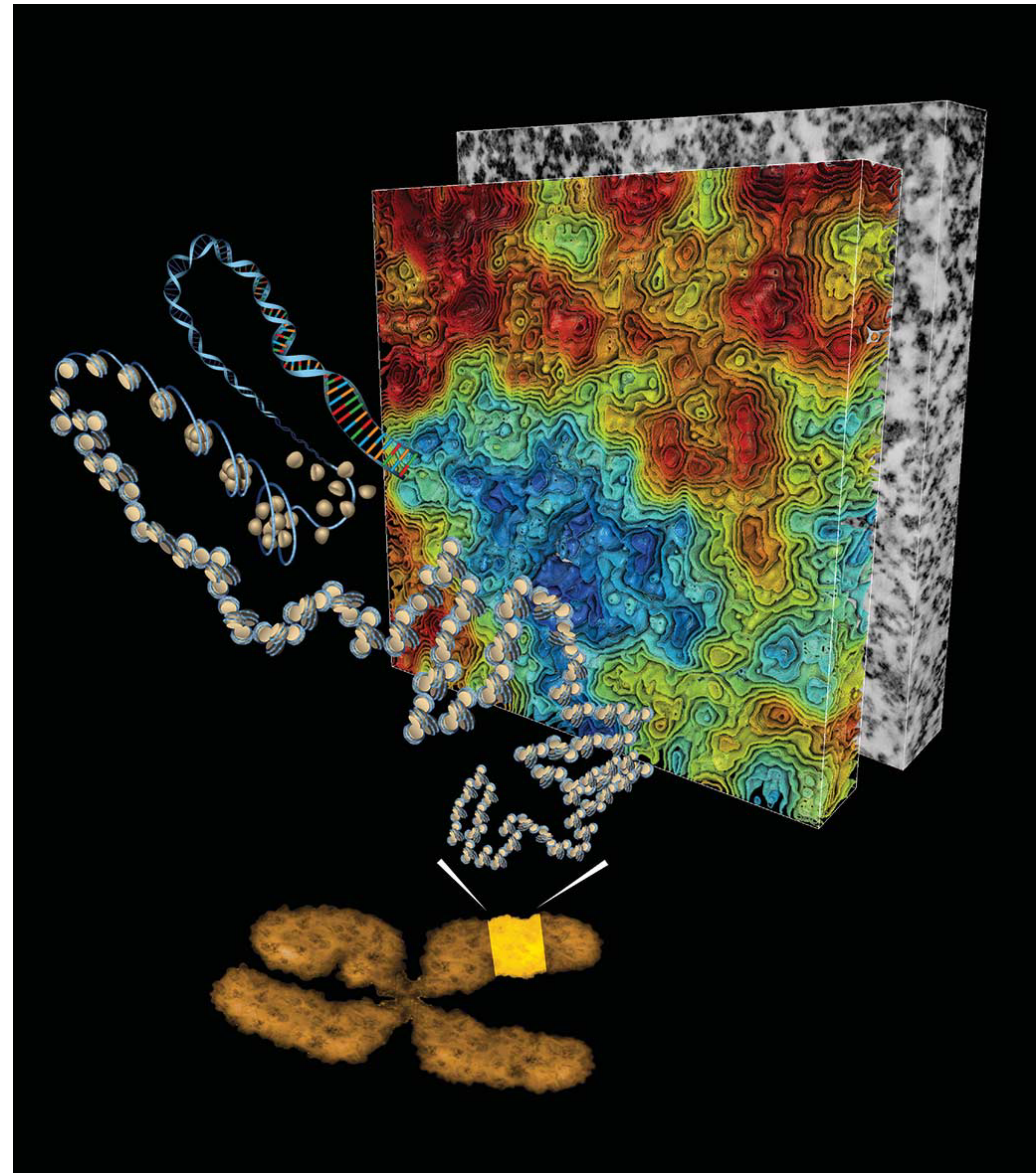
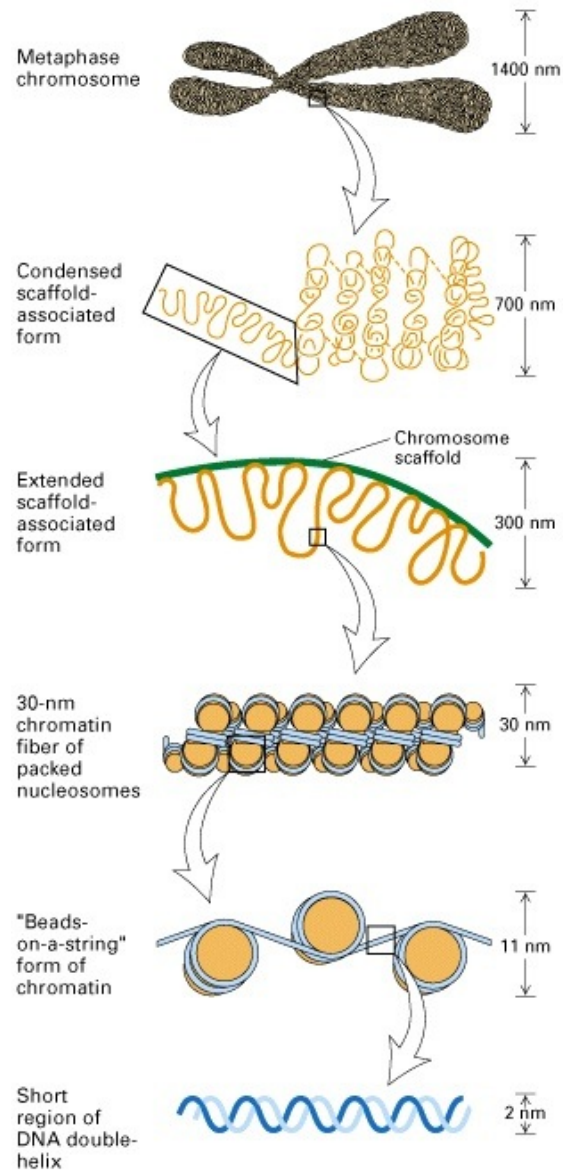
>80% of the genome is functional

ENCODE defined product or reproducible biochemical activity

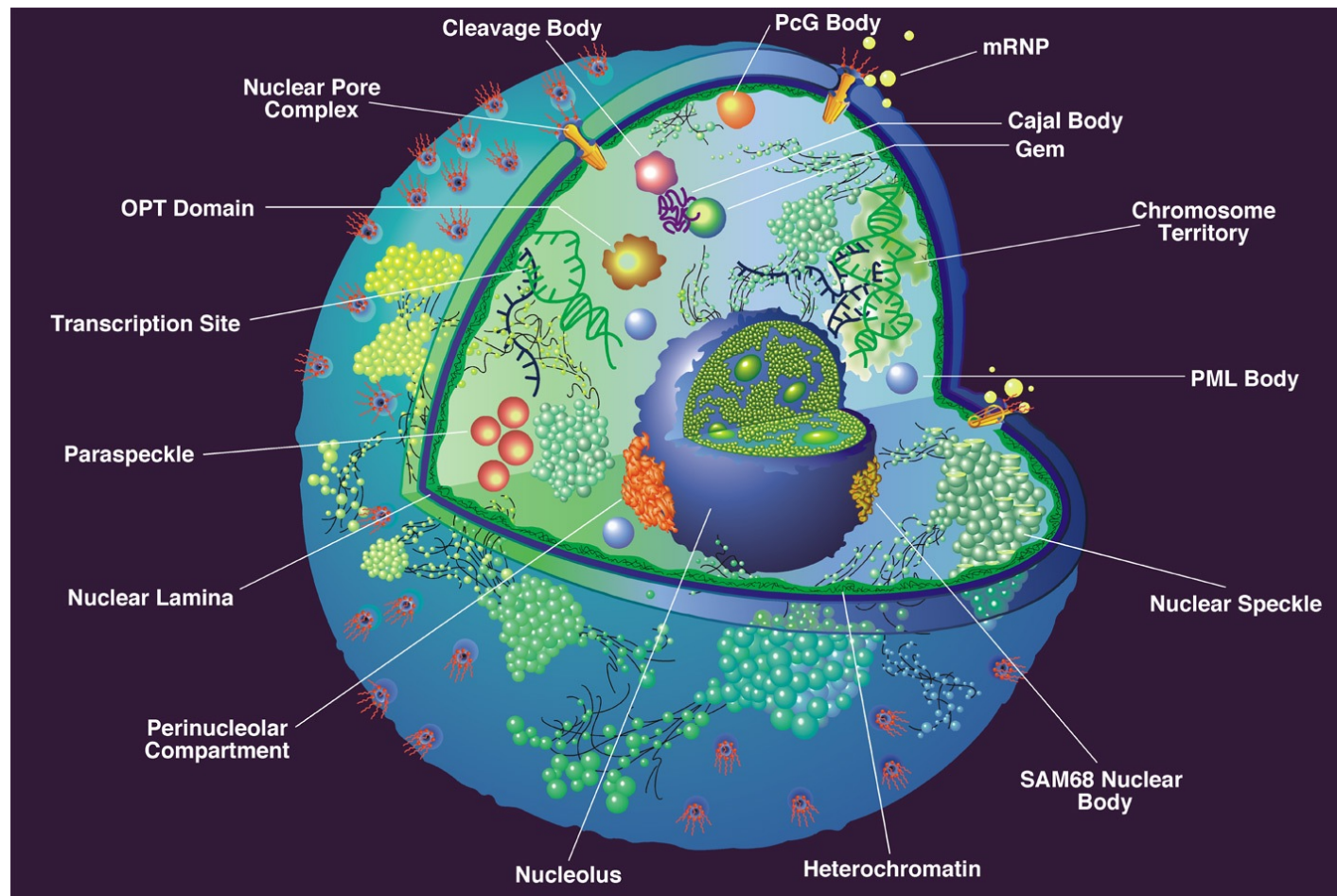
<http://www.nature.com/epigenomeroadmap>



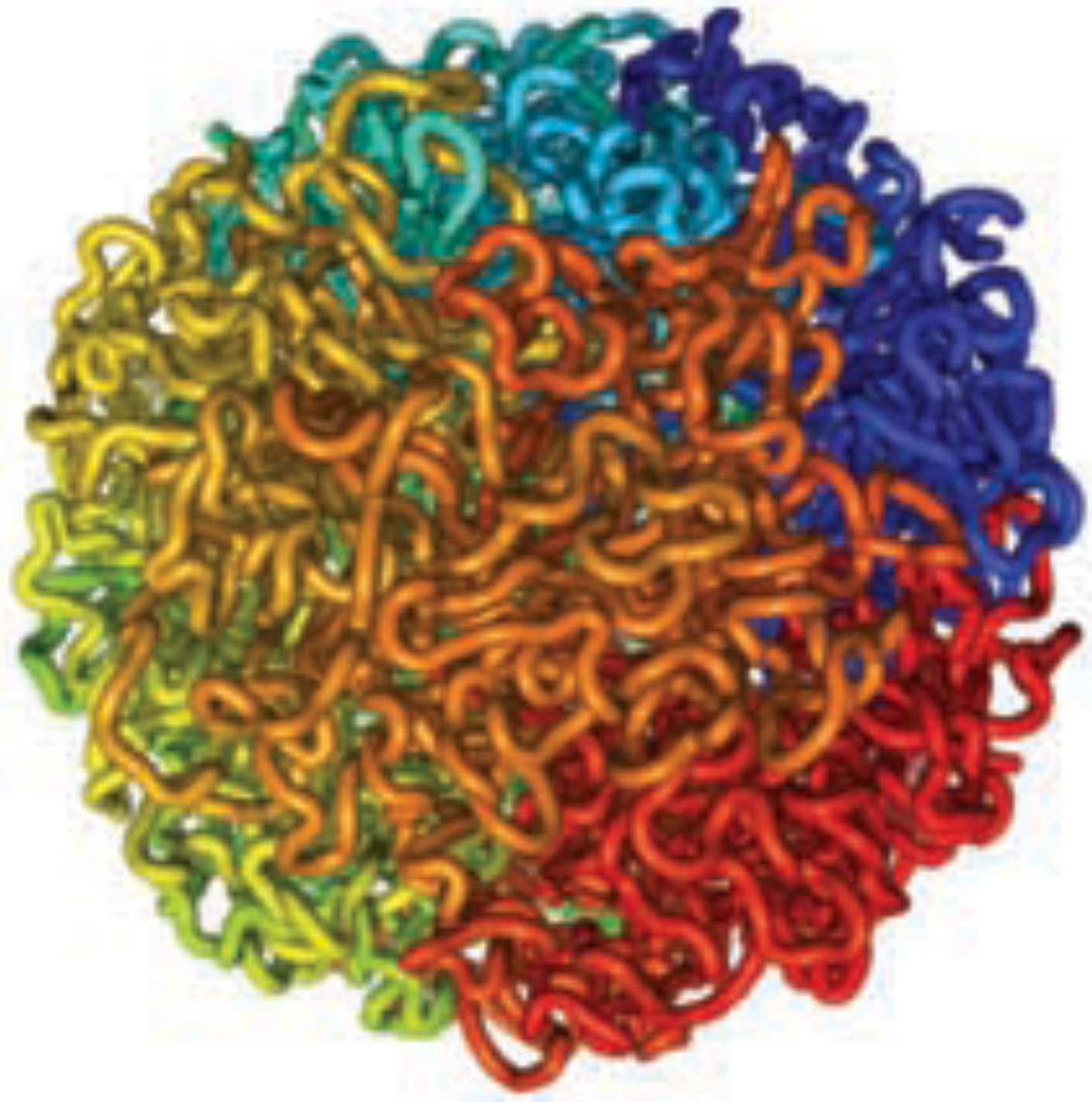
chromatin structure

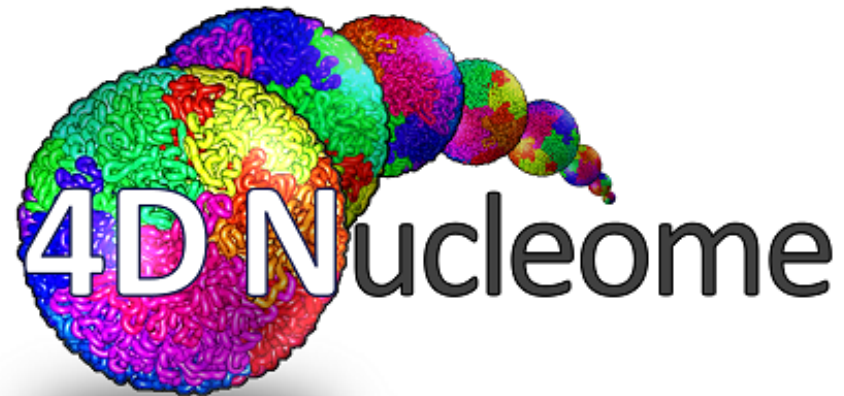


nuclear bodies



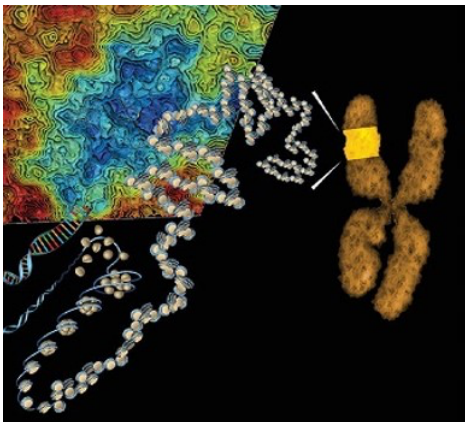






<https://commonfund.nih.gov/4dnucleome>

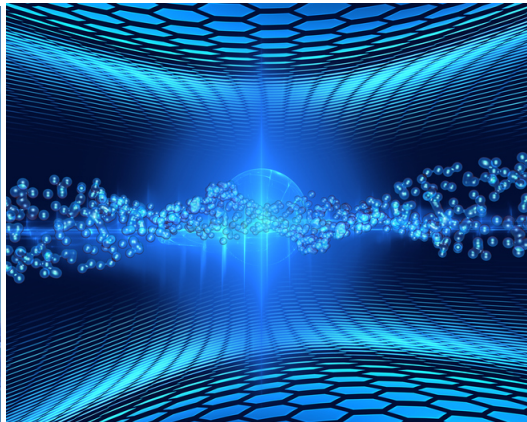
visualizing the nucleus in 3D



genes in motion



genome organization



genome organization in development

